Composition and mobilization of particulate organic matter from retrogressive thaw slump impacted streams

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I. BACKGROUND

The source and composition of organic matter can play an important role in metabolism in aquatic systems. Although thermokarst processes are thought to mobilize significant quantities of particulate organic matter (POM), and its organic carbon component (POC), to aquatic systems (1), we know little about its composition and fate, despite permafrost-origin dissolved organic carbon (DOC) having been shown to be easily degraded to CO_2 (2-4).

In the western Canadian Arctic, a warming and wetting climate has intensified thaw slumping (5). These features thaw large volumes of glacigenic sediments and can increase total suspended sediment flux in streams by orders of magnitude (6). While these high sediment concentrations have been shown to have negative impacts on stream biota (7) the alteration of organic matter pools, especially POM pools, and



Large thaw slumps, or those well connected to streams, increase organic carbon yields in streams by orders of magnitude, primarily driven by increases in POC. Impacted streams switch from being DOC dominated systems to POC dominated systems. This also reflects changes in the partitioning of organic matter pools.



IV. RESULTS



1e+3



Thaw slump runoff contains dissolved organic matter (DOM) richer in nutrients relative to DOM in un-impacted streams. However, DOM mobilized by thaw slumps does little to alter the DOM pool

Thaw slump runoff contains particulate organic

(A)

(B)

its impact on aquatic ecosystems has not been addressed.



Fig. 2. A) Total (TOC), particulate (POC) and dissolved (DOC) organic carbon instantaneous yield (mg s⁻¹ km⁻²) and B) POC to DOC ratio upstream and downstream of

Sediment



How do thaw slumps alter organic carbon **<u>delivery</u>** to streams and how does this effect differ in the dissolved vs. particulate phases?



How do thaw slumps alter the **composition** of organic matter in streams and how does this effect differ in the dissolved vs. particulate phase?



thaw slump sites. Error bars indicate standard error of the mean of samples obtained from June to August (n=2 HA-HD; n=3 SA – SE).



Impacted streams have an older PO¹⁴C age similar to slump runoff suggesting significant mobilization of permafrost POM

Decreasing thickness of permafrost thaw



upstream, from, and downstream of the runoff of three active thaw slumps. Error bars represent 1σ confidence limits.





SC Headwall



Fig. 4. C: P vs C: N of A) dissolved and B) particulate organic matter upstream, from, and downstream of slump runoff. Error bars indicate standard error of the mean of samples obtained from June to August (n=2 HA – HD; n=3 SA – SE). Note: Some sampling points for Fig 5A are missing due to dissolved phosphorus values being near the detection limit.

Questions/Comments?

I'm always happy to chat about my research!

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VI. CONCLUSIONS & FUTURE DIRECTIONS

Thaw slumps increase total organic carbon and organic matter delivery to streams by orders of magnitude. However, this increase in delivery is primarily due to increases in the particulate phase. Thaw-slump-mobilized POM is depleted in nutrients compared to POM present in un-impacted streams and may be a poorer quality food source to stream organisms. This, along with extreme loading of particulate material in streams, may have adverse impacts on stream biota both in adjacent aquatic systems and downstream environments.



Fig. 1. A) Thaw slump sites, Peel Plateau, NWT. B) Sampling schematic. Whole water samples were obtained upstream, from, and downstream of slump runoff. Samples were collected 1-2 times from each thaw slump from June to August in 2015.

Ongoing and Future Research Directions

Is POC mobilized by thaw slumps readily biodegradable? > How does the impact of thaw slumps differ in different watershed units?

> To what degree is POC mobilized by thaw slumps available for transport downstream and how does it alter watershed organic matter flux?

ACKNOWLEDGEMENTS

I would like to thank Joyce Kendon, Luke Gjini, and Scott Zolkos for their assistance both in the lab and during field sample collection. I would also like to acknowledge the provision of lemonade by Luke Gjini and Scott Zolkos at field site SA. Thank you to Cara Bulger for assistance with field sampling design. Further thanks to Scott Zolkos for assistance with field logistics prep and the never-short-supply of amazing field pictures. A huge thank you to wildlife monitors Christine Firth and Elizabeth Jerome for guidance to field sites and help during field sample collection and to the Tetlit Gwich'in Renewable Resources Council and Aurora Research Institute for their logistical support. This research was funded in part by the Aurora Research Institute, the Garfield Weston Foundation, the University of Alberta and UAlberta North, the Northern Scientific Training Program (NSTP), the Natural Sciences and Engineering Research Council (NSERC), and the Polar Continental Shelf Program (PCSP). Employee salary support from the Environment Canada Science Youth Horizons Internship program also helped support the employment of Joyce Kendon. Data in this poster was managed and reworked through the use of Microsoft Access. Thanks to Dr. Erin Bayne for providing a course on relational databases at the University of Alberta that has saved me both time and frustration. Thank you to Mingsheng Ma and BASL staff for lab support. Dr. Vincent St. Louis suggested presenting flux data as yield data.

Whole water samples and discharge measurements (current meter, area-velocity method) were obtained upstream, downstream, and from RTS outflows and from eight RTS sites (Fig. 1). Samples for dissolved analyses were allowed to settle for 24 hours with 10% HCI). Samples for dissolved analyses, and from eight RTS sites (Fig. 1). Samples for dissolved analyses were allowed to settle for 24 hours with 10% HCI). Samples for dissolved analyses were allowed to settle for 24 hours prior to filtration through a pre-combusted GF/F filter. For particulate analyses, and from eight RTS sites (Fig. 1). Samples were collected in HDPE bottles (pre-leached for 24 hours with 10% HCI). Samples for dissolved analyses were allowed to settle for 24 hours with 10% HCI). Samples for dissolved analyses were allowed to settle for 24 hours with 10% HCI). Samples for dissolved analyses were allowed to settle for 24 hours with 10% HCI). Samples were collected in HDPE bottles (pre-leached for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours with 10% HCI). Samples were allowed to settle for 24 hours a separate bottle was collected and shaken thoroughly before a known volume of sample water was filtered through pre-weighed and pre-combusted GF/F filter (diameter = 47mm) within 24 hours of collection. Filters were stored frozen until shipment to the U of A. Filter samples were dried at 60°C and weighed to determine total suspended sediment concentration (mg L⁻¹). Samples for POC were fumigated for 24 hours at 60°C with 12M HCI. Fumigated samples for POC were packed into tin capsules and analyzed by an elemental analyzed by an elemental analyzed by an elemental analyzed to a continuous flow isotope ratio mass spectrometer (IRMS, UC DAVIS Stable Isotope Facility) along with paired non-acidified samples for particulate nitrogen (PN) analysis. Particulate phosphorus samples were analyzed using the ash-hydrolysis method following (8). DOC concentration was determined as non-purgeable organic carbon (NPOC) from the best three of five injections, with a coefficient of variation <2% or similar, on a Shimadzu TOC-V analyzed for ammonia and nitrate/nitrite via colorimetry at the University of Alberta, Biogeochemical Analysis Laboratory (BASL) following modified versions of Standard Methods (SM) 4500-NH3-B,H and US EPA 353.2 respectively. Total dissolved nitrogen (TDN) samples were analyzed via colorimetry at BASL following SM 4500N-B. Soluble reactive phosphorus samples were analyzed via colorimetry at BASL following SM 4500N-B. Soluble reactive phosphorus samples were analyzed via colorimetry at BASL following SM 4500N-B. Soluble reactive phosphorus samples were analyzed via colorimetry at BASL following SM 4500N-B. Soluble reactive phosphorus samples were analyzed via colorimetry at BASL following SM 4500N-B. Soluble reactive phosphorus samples were analyzed via colorimetry at BASL following SM 4500N-B. Soluble reactive phosphorus samples were analyzed via colorimetry at BASL following S Institute laboratory. Total dissolved phosphorus (TDP) was analyzed via colorimetry at BASL following SM 4500 P-B. Dissolved pools (TDN and TDP respectively) minus their respective inorganic pools (DIN and SRP respectively). All calculations were made using molar concentrations. Watershed areas, delineated using Canadian CDED CDEM and NHN stream network datasets using ArcGIS, were combined with discharge and concentration data to calculated instantaneous fluxes (Fig. 2a).

Fumigated samples for PO¹⁴C were sent to A.E. Lalonde AMS Laboratory where radiocarbon analyses were performed on a 3MV accelerator mass spectrometer (AMS) by High Voltage Engineering. Measurements were normalized with respect to the reference material Oxalic II (F14C=1.34) and ages are calculated using the Libby 14C half-life of 5568 years. Samples for dissolved organic carbon (DOC) were measured on a Shimdazu TOC-V analyzer.

SUPPLEMENTARY METHODS

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